



OFFICE OF WATER PROGRAMS

BUREAU OF CLEAN WATER

TECHNICAL DEVELOPMENT OF A THERMAL FISH INDEX

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INTRODUCTION

This document is intended to describe the purpose, applicability, and development of a thermal fish index (TFI) that serves as a multidisciplinary tool for management centered around fish and their role in 25 Pa. Code § 93. Specifically, “Uses” are discussed to establish initial context for making assessments and evaluations, pursuant to Water Quality Standards, using fish. The technical development of a TFI is conducted and the general results are discussed to create an initial foundation for assessment and evaluations. Specific methods stemming from this technical document will be presented independently.

Table 1 in 25 Pa. Code § 93.3, lists five Use Categories as: Aquatic Life, Water Supply, Recreation and Fish Consumption, Special Protection and Other. Each of these categories is specifically intended to protect and support the resource and/or the user of the resource. Aquatic Life defines four sub-category Aquatic Life Uses (ALUs), three of which – Cold Water Fishes (CWF), Warm Water Fishes (WWF) and Trout Stocking (TSF) – are narrative definitions of biological communities along a thermal regime (Table 1). The intent of categorical ALU and sub-categorical definitions are to measure a waterbody’s ability to support the defined ecological communities (i.e., water quality and habitat are supportive), with numeric criteria established to be protective of aquatic life. To this end, development of biologically-based ALU assessments must be calibrated and responsive to changes in water quality and habitat as a reflection of waterbody condition. Alternatively, Recreational Use (RU) assessments are developed to protect and support the user of the waterbody (Table 1) with numeric RU criteria established to be protective of human health.

Maximum temperature criteria are provided for defined times of the year for each of these sub-categorical ALUs. Temperature criteria in § 93.7 are applied to heated waste sources regulated under 25 Pa. Code §§ 92a and 96. Temperature limits apply to other sources when they are needed to protect designated and existing uses. In other words, temperature criteria are applied to specific cases and are not used for broad assessments of ALU. As indicated from the ALU definitions, an appropriate thermal evaluation includes a biological assessment based on instream flora and fauna, with specific mention of fish species.

Table 1. 25Pa Code 93.3 Aquatic life and Recreation uses are listed as;

Aquatic Life	
<i>CWF - Cold Water Fishes</i>	Maintenance or propagation, or both, of fish species including the family Salmonidae and additional flora and fauna which are indigenous to a cold water habitat.
<i>WWF - Warm Water Fishes</i>	Maintenance and propagation of fish species and additional flora and fauna which are indigenous to a warm water habitat.
<i>MF - Migratory Fishes</i>	Passage, maintenance and propagation of anadromous and catadromous fishes and other fishes which move to or from flowing waters to complete their life cycle in other waters.
<i>TSF - Trout Stocking</i>	Maintenance of stocked trout from February 15 to July 31 and maintenance and propagation of fish species and additional flora and fauna which are indigenous to a warm water habitat.
Recreation	
<i>B – Boating</i>	Use of the water for power boating, sail boating, canoeing and rowing for recreational purposes when surface water flow or impoundment conditions allow.
<i>F – Fishing</i>	Use of the water for the legal taking of fish. For recreation or consumption.
<i>WC – Water Contact</i>	Use of the water for swimming and related activities.
<i>E – Esthetics</i>	Use of the water as an esthetic setting to recreational pursuits.

Freshwater fishes are important indicators of temperature as they are obligate poikilothermic, meaning their internal body temperatures are dictated by the ambient surrounding water temperature (Wood and McDonald 1997, Beitinger et al. 2000). Thermal preference and tolerance vary among species (Wehrly et al. 2003, Yoder 2006), creating unique assemblages of fishes along a continuous gradient, upstream to downstream. These longitudinal changes in fish assemblages parallel important shifts in loading, transport, and utilization of organic matter from headwaters to mouth that form the river continuum concept (RCC; Vannote et al. 1980). The thermal zonation of fishes along a longitudinal gradient has been realized for nearly a century (Carpenter and Huxley 1928) and biological zones have been identified based on the occurrence of dominant fishes as “indicator species” (Huet 1959).

The use of indicator species may be appropriate where only presence-absence data are available, but the use of indicator species in biological assessments tends to lack responsiveness to degradation along a continuous gradient (Fausch et al. 1990). For example, when an indicator species is absent due to stress, any additional stress on the system will have no measurable effect. An alternative to using indicator species is the use of all species in an assemblage and their relative abundance based on taxonomy, traits, and tolerance values to make biological assessments along a broad range of stress. The shift from indicator species to a more broad-scale, assemblage-based approach largely began in the 1970's-80's and the application of these concepts were first realized by the seminal introduction of the Index of Biotic Integrity (IBI) conceptualized by Karr (1981). As assemblage-based concepts began to evolve from indicator species concepts, regulatory definitions evolved as well. Historic ALU definitions were largely dependent on using trout species (family Salmonidae) as an indicator of a cold water community, as evident from the definitions from the late 1960's (Table 2), to what they are today (Table 1).

It is important to note that sub-categorical ALU definitions that make specific mention of trout, use trout as an indicator of natural thermal communities; where trout populations are being completely supported indicates that additional flora and fauna indigenous to a cold water habitat may also be supported (CWF). Where trout are not supported by waterbody conditions indicates that additional flora and fauna indigenous to a warm water habitat may be supported (WWF). Where trout are partially supported indicates an ecological community intermediate of CWF and WWF exists that is not completely described by the sub-categorical definition of TSF. While trout have excellent socioeconomic value, the socioeconomic value of trout cannot be included in ALU, as it is appropriately included and protected under the sub-categorical Fishing Use, within RU (Table 1).

Table 2. Historic sub-categorical aquatic life uses obtained from Article 301 of the Sanitary Water Board Rules and Regulations, Commonwealth of Pennsylvania, Water Quality Criteria (1968).

<i>Cold Water Fishes</i>	Maintenance or propagation, or both, of fish species of the family Salmonidae and fish food organisms.
<i>Warm Water Fishes</i>	Maintenance and propagation of fish food organisms and all families of fishes except Salmonidae.
<i>Migratory Fishes</i>	Passage, maintenance and propagation of anadromous and catadromous fishes and other fishes which ascend to flowing waters to complete their life cycle.
<i>Trout (Stocking Only) *</i>	Warm water fishes and trout stocking

* Added December 20, 1967.

On the surface, sub-categorical ALU definitions could be interpreted as a form of thermal assessment as CWF, TSF and WWF are considered hierarchical along a thermal gradient. However, these definitions have inherent complexities that present challenges for assessment purposes. Specifically, streams under natural (or near natural) conditions may not always support CWF (e.g., large streams, rivers). Additionally, the interpretation of the ALU definitions have traditionally relied heavily on the presence of fish (trout in CWF) to fulfil the “maintenance” requirements, and the presence of young-of-year or multiple age-classes of fish (trout in CWF) to fulfil the “propagation” requirements. This interpretation can be successfully applied to CWF when trout are present in high numbers but becomes more challenging as trout numbers are reduced (e.g., how many trout are needed to satisfy the narrative definition?). In other words, numerical thresholds may help alleviate subjectivity while providing consistent interpretation of narrative definitions. Furthermore, species within the trout family (Salmonidae), need to clearly demonstrate a positive response to good water quality, if their use as a potential indicator of waterbody condition is to be meaningful. Preliminary investigations of trout density and abundance as an indicator of water quality suggests responses to water quality may be variable overall and species-specific (Figure 1).

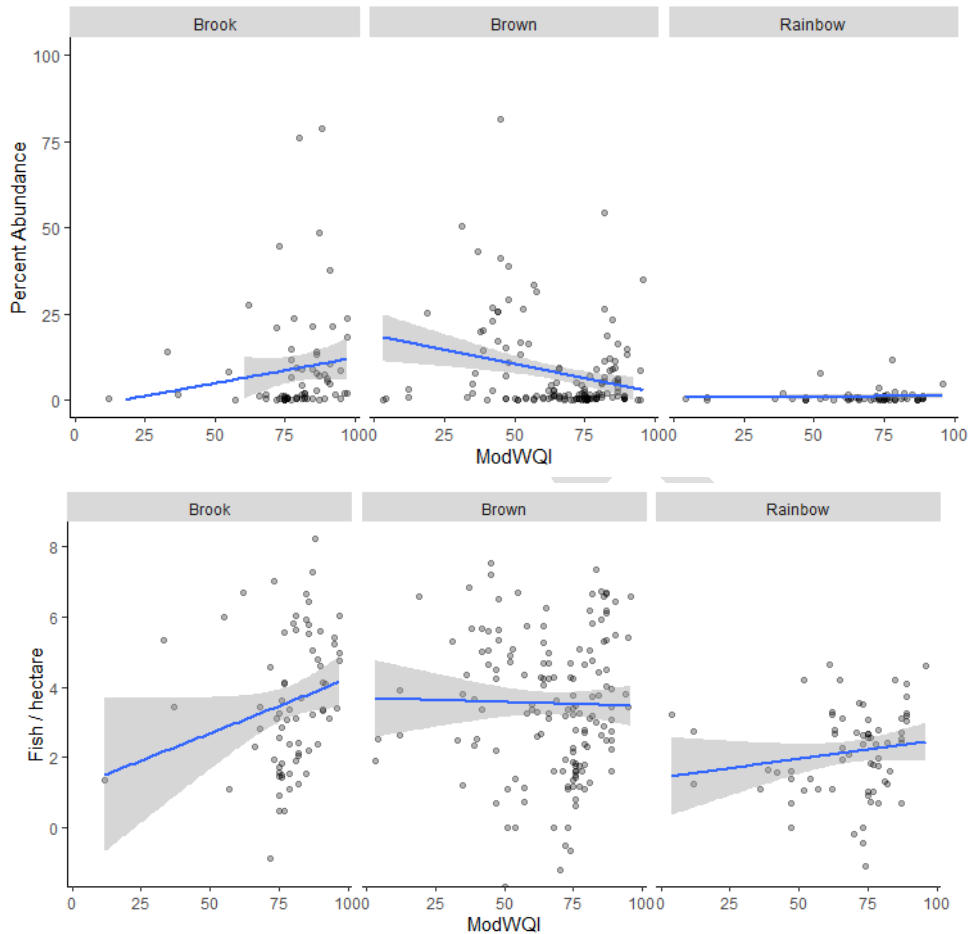


Figure 1. Three common trout species found in Pennsylvania, percent abundance and density (log transformed) as the number of fish per hectare, in response to water quality. Water quality was measured with the modified water quality index (ModWQI), at all sites where present. The ModWQI measures water quality stress from poor to good, on a continuous scale from 0-100, respectively.

Interpretation of ALU narrative definitions can be bolstered through the development of numeric thermal assemblage classes. This provides a shift from qualitative implementation to a more quantitative description of fish assemblages, along a thermal gradient. Numerically describing the transition from “additional flora and fauna which are indigenous to a cold water habitat” to “additional flora and fauna which are indigenous to a warm water habitat” is possible by quantifying the transition of assemblages dominated by cold water species to assemblages dominated by warm water species. This transition is considered continuous in nature as opposed to binary, meaning there will be assemblages dominated by warm water species that may still have cold water species present. This transitional or “cool water” assemblage appears to align with the TSF use interpretation presented, where stocked trout may be seasonally maintained within a warm water assemblage. However, important differences are noted between

the TSF use and a transitional assemblage that may preclude quantification of TSF directly. Herein, a transitional assemblage is considered a segue of biological assemblages intermediate of cold and warm assemblages, based on environmental changes along a waterbody's continuum (e.g., longitude, slope, temperature). Subsequently, biological assessments should be directed towards quantifying a transitional assemblage (as opposed to TSF), as a measure of a waterbody's ability to support this natural assemblage. To avoid confusion between established uses in Chapter 93 and natural assemblages, the terms cold water assemblage (CWA), transitional assemblage (TSA) and warm water assemblage (WWA) will be used hereafter to describe thermal assemblages from an ALU assessment perspective. These terms are used to describe assemblage classes of fishes along a thermal (and environmental condition) gradient; and should in no way be considered redefinitions of ALUs.

As previously stated, ALU assessment tools are designed to evaluate a waterbody's condition by measuring changes in biological assemblages, in response to stress. The natural thermal zonation of fishes along a longitudinal gradient can be altered by anthropogenic stressors (Caissie 2006, Stanfield and Kilgour 2013) that include but are not limited to: deforestation (Brown and Krygier 1970, Jones et. al 1999, Burcher et. al. 2008), urbanization (Brown et. al. 2005, Nelson and Palmer 2007), groundwater manipulation (Poole and Berman 2001, O'Driscoll and DeWalle 2006), impounding (Ward and Stanford 1983, Lessard and Hayes 2002), thermal effluents (Coutant 1975, Shuter et al. 1980), and global climate change (Eaton and Scheller 1996, Mohseni et al. 2003, Nelson and Palmer 2007). Thermal regimes can also be affected by natural factors that may combine to shape the fish assemblages found within the waterbody. Common natural effects that influence the thermal regime include measures related to latitude, elevation, slope, velocity, groundwater and canopy cover, among others. Less common effects may include measures of turbidity, basin orientation, or substrate characteristics. Effects of anthropogenic and/or natural factors are usually spatiotemporally stochastic and are both responsible (at varying degrees), in the formation of modern-day fish assemblages. This theory forms a physical habitat template and suggests that recovery from disturbances and the response of fish assemblages may vary accordingly (Southwood 1977, Poff and Ward 1990). In other words, as anthropogenic stress increases in a waterbody the natural thermal assemblage may adjust accordingly (Figure 2). Alternatively, as anthropogenic stress is mitigated (naturally or through management), the thermal assemblage may adjust accordingly. Therefore, it is important to note that the thermal response of fish assemblages is not exclusively limited to changes in temperature.

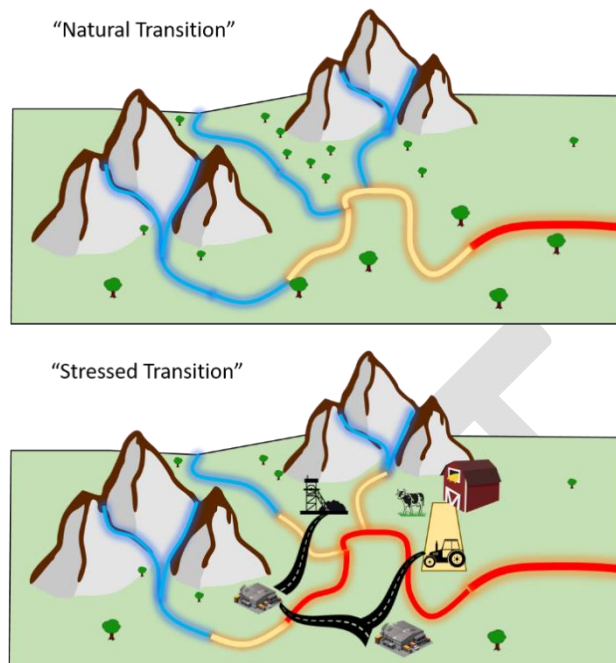


Figure 2. Theoretical example of natural longitudinal transition areas versus stress induced fish assemblage transitions. With applied stress to a cold water assemblage (CWA; blue), the CWA reduces, the transitional assemblage (TSA; yellow) is shifted upstream and the warm water assemblage (WWA; red) is expanded.

Historically, DEP has relied on bioassessments based on macroinvertebrate assemblages to make categorical ALU assessment determinations. Fish-based bioassessment tools offer a suite of benefits that compliment traditional macroinvertebrate-based assessments. Benefits include: 1) fish life-cycles are longer than macroinvertebrates, providing insight into acute and chronic exposure through time, 2) fish often respond to stress at different landscape scales than macroinvertebrates (Lammert and Allan 1999), 3) fish life history and tolerance information is widely available, 4) fish are relatively easy to identify, 5) fish have extensive socioeconomic value, and 6) fish provide important evidence of sub-categorical ALU narrative. Fish-based bioassessment tools are typically considered more complex than macroinvertebrate-based tools, in that: 1) fish can be highly mobile within dendritic freshwater systems 2) species distributions are based on zoogeographic factors that can make inter-basin comparisons challenging, and 3) barriers (physical and/or chemical) to recolonization efforts may delay recovery. Pennsylvania has had extensive zoogeographic influences that have shaped six major drainage basins and unique fauna within. Nearly 200 fish species, represented by 28 families, have been recorded from Pennsylvania's non-tidal waters (Stauffer et al. 2016). To overcome distributional challenges and have a fish-based bioassessment tool that can be broadly applied throughout Pennsylvania, focus should be shifted away from taxonomic

assessments (species level) and directed towards tolerance/preferences at the assemblage level. Additionally, focus should be directed towards relative abundance changes, as a way to help mitigate potential barrier-effects for recolonization. Specifically, this assessment development is directed towards thermal tolerances (or preference) of fish assemblages to make categorical ALU assessments, while numerically aligning assemblages with the intent of sub-categorical definitions, to the extent possible.

Although there is a great deal of literature concerning the thermal response of fishes, there is little information regarding the quantification of entire fish assemblages along a thermal gradient (but see; Zorn et al. 2002). The following represents the introduction of a metric (the TFI) that quantifies entire assemblages' thermal preference as a numerical description of how "cold" or "warm" a fish assemblage is, based on a unitless scale. The TFI ranks assemblages from coldest to warmest along a 2.0 to 10.0 scoring gradient, respectively.

METHODS

Index Calculation

Fish species were designated within a thermal class as determined from thermal studies compiled by Eaton and Scheller (1996) and Lyons et al. (2009). Eaton and Scheller (1996) ranked each species on a three-tiered classification of Cold, Cool, and Warm from streams across the continental U.S., whereas Lyons et al. (2009) utilized an additional fourth tier by splitting Cool into Cool-transitional and Warm-transitional in Wisconsin and Michigan streams. Tiered delineations were converted to five-tiers with associated numerical values: 1-Cold (Cd), 2-Cold-Cool (CdCl), 3-Cool (Cl), 4-Cool-Warm (ClWm), and 5-Warm (Wm), similar to Coker et al. (2001), to normalize any disagreement between delineations. The list of Pennsylvania fish taxa and their thermal delineations were then independently sent to regional experts familiar with fishes of the Northeastern and Mid-Atlantic U.S. to delineate taxa not directly addressed by Eaton and Scheller (1996) and Lyons et al. (2009). Final delineations from regional experts were chosen based on modal values (Appendix A). Where modal values were not achieved, the delineations were made by using arithmetic mean rounded up or down by considering latitudinal distributions and habitat preferences for each species, similar to Coker et al. (2001).

To calculate the TFI the number of individuals within each thermal class, as a percentage (e.g., 20% cold water individuals), was calculated. A weighted average was obtained by multiplying the numeric value for the thermal class by the percent of individuals, summed across classes. The final value is then multiplied by two to expand

and standardize the range from two to ten, coldest to warmest respectively (Table 3). Calculation of the TFI follows;

EQUATION

$$TFI = \left(\sum_{1}^{5} NP_i \right) 2$$

Where *N* is the numeric value for the thermal class and *P* is the percent of individuals at the *i*th thermal class, summed across all five classes and multiplied by two.

Table 3. Example of proportional abundance shifts of individuals within a thermal class, across the five thermal classes, and the resulting thermal fish index (TFI) score.

Cold 1	Cold-Cool 2	Cool 3	Cool-Warm 4	Warm 5	TFI Score
1.00					2
0.60	0.30	0.10			3
	0.60	0.30	0.10		5
		0.60	0.30	0.10	7
		0.10	0.30	0.60	9
				1.00	10

Reference Condition and Stressor Gradient

A least-disturbed (LD) approach was used to develop reference condition based on Stoddard et al. (2006), or the “best available” condition. The criteria for establishing LD condition was determined *a priori* and applied consistently across streams of all sizes to allow for assemblage characterization, along a longitudinal gradient. Three major stress categories were identified as stressed (S), moderately stressed (M) and LD (Figure 3). Two major stressors on aquatic environments were used to delineate stress categories: water quality and habitat. Water quality stress was measured using a modified version of DEP’s water quality index (WQI), originally described by Wertz and Shank (2019). The original WQI used 21 parameters to inform stress condition along a land-use-similarity index (range = 0-100, S to LD, respectively). The modified WQI (modWQI) was reduced to 18 parameters (Table 4), which increased the number of fish sites available with paired water quality. Instream habitat measures were conducted following a modified version of the U.S. Environmental Protection Agency’s (USEPA’s) Rapid Bioassessment Protocols for Use in Streams and Wadeable Rivers (RBP III) (Plafkin et al. 1989, Barbour et al. 1999) associated with DEP and Susquehanna River Basin Commission (SRBC) collection methods at each fish site (Shull and Lookenbill 2018,

Shank et al. 2016). Habitat measures were standardized into a habitat category score (Habcat; range = 1-4, LD to S, respectively) based on available measures of: sedimentation, embeddedness, sand, silt and detritus, as measures of habitat condition varied across sampling methods (i.e. wadeable vs. nonwadeable). Finally, a dam proximity criterion was added to ensure fish sampling sites were not close to habitat-modified systems, or barriers to migration, that could potentially influence the fish assemblage (Table 5). Dams and impoundments were identified on a site-specific basis to better identify likely impacts (e.g., small impoundments are more likely to influence small streams than large rivers).

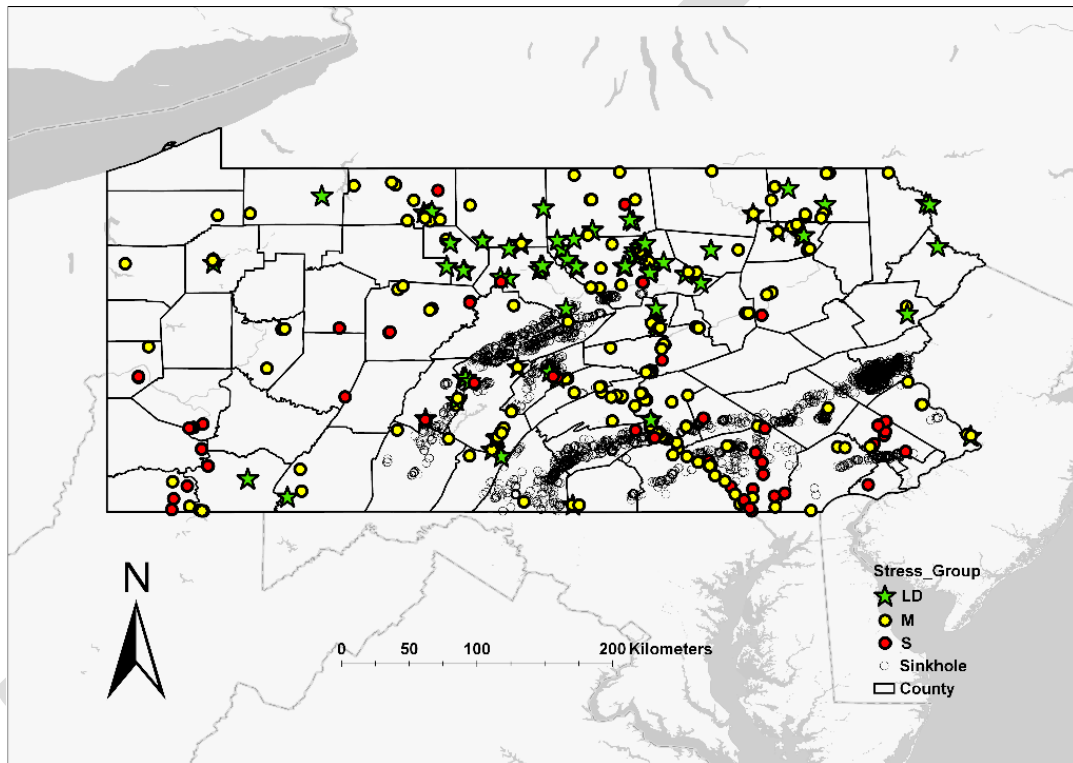


Figure 3. Map of sites considered least disturbed (LD), moderately stressed (M), and stressed (S) across a gradient of water quality and habitat conditions. Open circles represent sinkhole locations and relative density.

Table 4. Water quality parameters (n=18) used to create a modified water quality index (modWQI).

PARAMETER
ALKALINITY, TOTAL
ALUMINUM, TOTAL
AMMONIA TOTAL AS NITROGEN
BROMIDE, TOTAL
CALCIUM, TOTAL
CHLORIDE, TOTAL
DISSOLVED SOLIDS, TOTAL
HARDNESS, TOTAL
IRON, TOTAL
MAGNESIUM, TOTAL
MANGANESE, TOTAL
NICKEL, TOTAL
pH
PHOSPHOROUS, TOTAL
SPECIFIC CONDUCTIVITY @ 25.0 C
SULFATE
SUSPENDED SOLIDS, TOTAL
ZINC, TOTAL

Table 5. Least-disturbed and stressed criteria based on water quality using a modified water quality index (modWQI), habitat from a categorical measure (Habcat) and dam proximity.

Description	Freestone		Limestone	
	Least Disturbed Criteria	Stressed Criteria	Least Disturbed Criteria	Stressed Criteria
modWQI score	> 60	< 40	> 40	< 20
Habcat score	1	3 or 4	1 or 2	3 or 4
Proximity to dam or impoundment	> 1.5 km		> 1.5 km	

Datasets

Prior to development, water quality and habitat data were spatiotemporally paired with fish assemblage data; the full dataset was then inspected for potential outliers and anomalies. Three potential issues were considered at this stage and addressed accordingly. First, if an assemblage had >10% of the individuals not identified to the species-level (e.g., not having a thermal class), the TFI score was considered not representative of the assemblage and the site was removed. Second, samples with less than 50 individuals were investigated for potential cause and representativeness. Potential causes for low sample sizes were investigated for: 1) appropriate application of collection protocols (e.g., electrofishing settings, survey distance and time), 2) potentially toxic water quality conditions, and 3) near sterile conditions (e.g., extremely low productivity). Only two of these causes were identified in the dataset. Toxic conditions were observed at sites with extreme acid mine drainage and near sterile conditions were observed in extreme headwaters. In both cases all samples were still considered to be representative of the overall site conditions and were retained. Lastly, sites were coded based on spatiotemporal representation of at least one water chemistry sample to fish sample location and time. Sites were coded from zero to three, best to worst expected representation, respectively. All three's (those most-likely to be unrepresentative) were removed from the dataset.

The full dataset was divided into three subsets; 1) precision dataset, 2) calibration dataset, and 3) validation dataset. The precision dataset was first partitioned from the full dataset to reduce any pseudo-replication or spatial-autocorrelation issues (Sokal and Oden 1978, Hurlbert 1984). Since the stress measures used have a strong temporal component (i.e. based on instream measures instead of land use) sample independence was defined in an attempt to retain repeated samples from the same site that have measurable, spatiotemporal change in water quality or habitat. Independent samples were identified, randomly across samples at the same site, as having either a five-point modWQI score change or a one-point change in Habcat score. These respective temporal changes to water quality or habitat are biologically meaningful as they can occur as a result of anthropogenic activities, which would be expected to influence fish assemblages through time. Samples that didn't meet this definition were regarded as repeat measures and were used in the precision dataset (samples removed; $n= 193$). The temporal strength of this method negated the need for temporal precision estimates as it treated all duplicate and replicate samples, given similar water quality and habitat conditions, the same. In other words, repeated sites were considered standardized by habitat and water quality, to ensure variability was associated with natural conditions (i.e. seasonal, sampling, processing). This result was desired as "duplicating" fish sites presents theoretical challenges and is best considered "replicating". The full development dataset included sites from all stress groups, was

then randomly split into a calibration and validation dataset (80/20, respectively; $n=360/90$). The LD sites within the calibration dataset were used for site classification purposes. The calibration dataset was used for development and the validation dataset served as an independent measure of performance.

Landscape Variables

Landscape variables were compiled at the local (stream segment) and watershed (total upstream catchment) scale. Local variables were obtained from the Appalachian Landscape Conservation Cooperative (AppLCC) stream classification system. The AppLCC contained data across six major variable types; size, gradient, temperature, hydrology, buffering capacity and confinement (Olivero et al. 2015). Of the six AppLCC variable groups available, temperature was the only variable group not used as it was based on fish assemblage data and was considered redundant. Catchment data was obtained by delineating upstream drainage areas for each site and measuring; area (km^2), density of sinkholes ($\#/\text{km}^2$) and the percent of limestone geology within the catchment. Sinkholes and limestone geology were specifically chosen to address potential relationships identified from previous studies relating to limestone and karst systems, and their effect on fish assemblages (Steffy and Kilham 2006, Carline et al. 2011, Kollaus and Bonner 2012). All landscape variables were compiled using ArcGIS Pro version 2.2 (ESRI 2018).

Site Classification

Boosted regression trees (BRTs) were used to classify LD and calibration datasets. Regression trees are a form of classification tree that utilize machine learning, where “boosting” generally improves performance from traditional regression trees by fitting multiple ‘simple’ models with an error term to avoid overfitting. Boosted regression trees, used in a continuous regression situation, use recursive partitioning to split data into homogenous groups and sub-groups based on between-group sum-of-squares, similar to analysis of variance (ANOVA), (Qian 2016, Elith et al. 2008). Regression trees (with or without boosting) have been utilized extensively for environmental modeling (Prasad et al. 2006, Breiman et al. 1984, Cutler et al. 2007, De'ath and Fabricius 2000, De'ath 2007) and groundwater studies (Trauth and Xanthopoulos 1997, Naghibi et al. 2016). All statistics were performed using R (R core team, 2016), BRTs were performed using package ‘rpart’ (Therneau and Atkinson, 2019), method = ANOVA for continuous response variable. Least disturbed sites were modeled to determine appropriate class for natural variables and the mean TFI within the predicted group for evidence of directional change. Results were analyzed for ecological relevance and minimal cross-validation error (Qian 2016). Similarly, BRTs were investigated in the calibration dataset to explore potential effects that may be problematic for analysis. Potential problematic issues may arise from stressors or site classification groups not represented in the LD

dataset. The results of the LD BRT classification schema were then applied to the calibration dataset and adjusted as needed to obtain final classification groups based on ecologically relevant concepts (i.e. RCC).

During preliminary data exploration investigations within the calibration dataset, using BRTs, the effect of karst geology became apparent (using sinkhole density within upstream catchment as a surrogate). This finding was in concordance with previous studies conducted in watersheds dominated by limestone geology, generally these systems have a unique ability to maintain cold water assemblages at increased stress levels, relative to their freestone counterparts (Steffy and Kilham 2006, Carline et al. 2011). Similarly, sites with increased sinkhole densities were observed to have reduced modWQI scores while still maintaining lower TFI scores, when compared to the rest of the dataset. Sites with ≥ 0.03 sinkholes/km² were identified from sites with < 0.03 sinkholes/km² in the full dataset, hereafter referred to as limestone (LS) and freestone (FS) stream types respectively. It is important to note that no LS streams met LD criteria for water quality as LS streams typically are found in wide, fertile valleys that tend to be dominated by agricultural practices. Habitat quality was also reduced in the limestone group, as many of these streams are typically lower gradient with moderate sand and gravel substrates. Additionally, the effect of habitat quality on the TFI score was apparent within the limestone group. Since this group of streams did not meet LD criteria for the freestone group the criteria was adjusted to accommodate (Table 5), after investigating the sites across their range of water quality. It is important to note that LS stress criteria were only adjusted due to lower (colder) TFI scores than FS streams under similar stress conditions. This is hereafter referred to as the “karst effect”. The karst effect began to dissipate after reaching significant size (~1,000 km² from this dataset) where TFI scores began to resemble that of similar-sized FS streams. To compensate for the karst effect, streams with sinkhole densities >0.03 /km² that were in catchments $>1,000$ km² were considered FS streams.

Data Analysis

Thermal fish index scores were investigated within the FS and LS datasets independently to determine thresholds that best align with ALU definitions, based on trout responses. Therefore, TFI scores were consolidated to integers (2=2-3, 3=3-4, ... 9=9-10) and the presence of trout within each integer group, as percent occurrence (PO), was used to determine the probability of trout occurrence. It should be noted that this was conducted at the sample level, as opposed to site level. Sample level is an under-estimate of the probability of presence at the site or stream level, while still providing confidence in sample probability. For example, if trout are present in low numbers in a waterbody the probability of capture increases with sample size. Furthermore, the percent abundance (PA) of trout averaged across TFI groups was calculated for comparison. This method produced four measures of trout response

along the TFI gradient; FS - (PO/PA) and LS - (PO/PA). These four trends were investigated to find inflection points (reduction of trout PA/PO), to establish TFI thresholds that best-represent a quantifiable transition from CWA to WWA, based on the TFI. The four measures were chosen to demonstrate the drastic difference across measures of occurrence and abundance. The measure of occurrence is very different than abundance, as trout can be present throughout a wide range of stream types at low abundance (e.g., one individual). Subsequently, the strict presence-of-trout measure reduces ecological meaningfulness without associated abundance measures.

Applying the natural classification schema from the LD to the calibration dataset allowed for measures of the TFI response to stress effects. Thermal scores were plotted and regressed to test for responsiveness to longitudinal gradient and stress levels. The datasets were tested for within-group normalcy and homogeneity of variance by inspecting residual distributions from linear models and Shapiro-Wilk tests. Final group sample sizes were relatively small and non-normal distributions were not all successfully transformed to meet parametric assumptions. Subsequently, Kruskal-Wallis chi-squared tests were used to measure among-group longitudinal differences of the final classification groups, using LD sites, followed by Dunn's test of multiple comparison, post hoc ($\alpha = 0.05$) adjusted using Bonferroni correction. Least disturbed sites were used to test for longitudinal response, minimizing effects from potential stressors. This procedure was repeated within-group based on stress level to measure significant differences in stress effect. Discrimination efficiency (DE) between LD and S sites was calculated to measure the TFI's ability to characterize stress (i.e. how much overlap exists between the LD and S TFI scores) (Barbour et al. 1999, Gerritsen et al. 2000). To measure DE, the percentage of stressed sites under the 75th percentile for LD sites was calculated by;

$$\%DE = \left(\frac{A}{B}\right) * 100$$

Where; A is the number of stressed sites scoring below the 75th percentile for LD range and B is the total number of Stressed sites.

After calculating DE, the 95th percentile of the LD sites within each group was used to establish impairment thresholds for assessment decisions. The 95th percentile is considered a high threshold for impairment which has two important considerations on assessments: 1) confidence in impairing a stressed site is increased, 2) confidence in not-impairing a stressed site is reduced. For example, if a stressed site is below the 95th percentile of the LD TFI range, it would be considered attaining. The decision to use the 95th percentile of LD sites is based on two reasons; 1) the modWQI is continuous in nature and allows for comparisons of stress response along a robust gradient of water quality across all stream classes, 2) using Habcat scores (1-4) there is more confidence

that LD sites are characterized as a 1 and less confidence that moderately stressed sites are characterized as a 4. For example, a moderately affected site with sedimentation issues is more likely to be classified as a 3 (stressed) or 4 (very stressed) than a 1 (not stressed); however, a site with severe habitat modifications (impoundment) may also be classified as a 3 or 4. Therefore, more confidence is placed on LD sites being accurately characterized, and less confidence on stressed sites being accurately characterized

Once impairment thresholds were established, validation was conducted to measure the assessments ability to classify sites not used in the development. Classification efficiency (CE) was calculated to measure the percentage of sites correctly classified based on the exceedance of established thresholds. The validation dataset was used to classify both impaired and attaining samples based on exceedance of impairment thresholds, measuring the percent correctly reclassified (i.e. the percentage of stressed sites being reclassified as impaired and the percentage of unstressed sites being reclassified as attaining).

The TFI metric was considered novel in both concept and application. A comparative analysis that demonstrated how the TFI compares to traditional metrics was needed to enhance understanding of metric function; both in ecological relevance and performance. Traditional metrics were calculated for the biological condition gradient level five (BCG; Davies and Jackson 2006), percent tolerant individuals, and percent omnivorous individuals. The BCG level five attribute is generally based on relative tolerance value of a species but also includes native/non-native status. A pairwise comparison using Spearman's rank correlation coefficient was conducted across metrics as well as the modWQI and Habcat to compare metrics responses to stress.

RESULTS

Thermal Assemblage Classes

The inflection point for trout PO was between TFI scores 6-7. Trout PA sharply decreased with TFI scores > 4 and the range of inflection was strongly noted between TFI scores 4-7 (Figure 4). Overall, the range of TFI scores from 5.0-7.0 indicates a strong transition in assemblages based on both trout abundance and occurrence. Upper thresholds were established to numerically define thermal assemblage classes that best represent the transition from an assemblage dominated by cold water species (TFI = 5.0), to dominated by cool water species (TFI = 7.0) and dominated by warm water species (TFI > 7.0), (Figure 4).

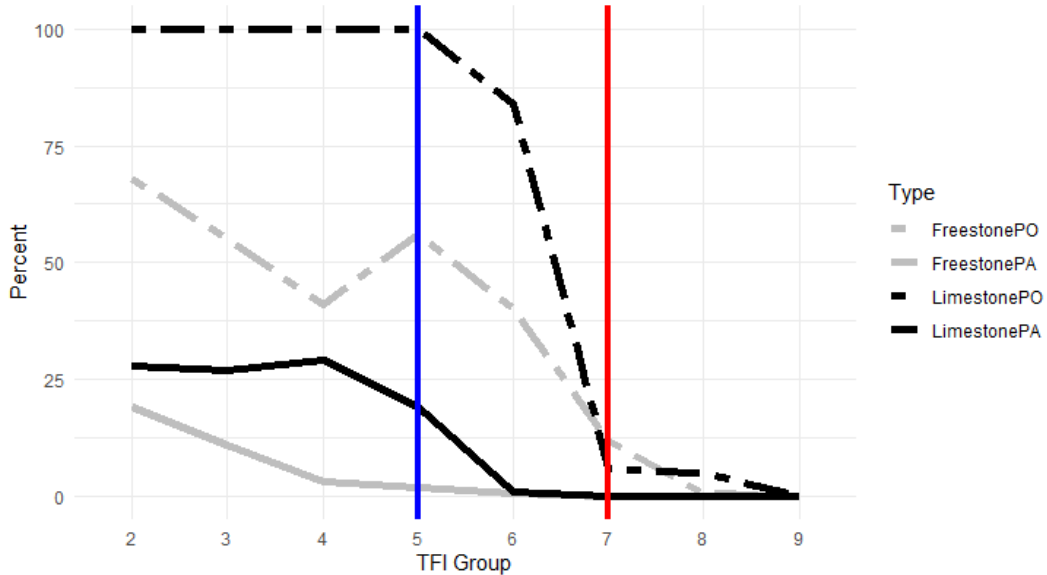


Figure 4. Percent abundance (PA) and occurrence (PO) of trout in both freestone and limestone streams. Dotted lines represent occurrence and solid lines represent abundance. The transition from cold water assemblage (CWA) to a warm water assemblage (WWA) is represented by blue and red vertical lines, respectively.

Modeled Results

Results from the BRTs using LD sites in FS streams indicated a strong longitudinal and slope effect, with minimal ecoregional effects in small catchments (Figure 5). Variable importance was partitioned relating to: stream size (41%), slope (32%), water quality (25%) and ecoregion (2%). Boosted regression trees in the LS dataset were conducted across all stress groups, as sample sizes from the LD sites precluded analysis. Boosted regression trees in LS streams also indicated a longitudinal and slope effect with additional karst effect (Figure 6). Variable importance was partitioned relating to: stream size (57%), karst and limestone geology (sum = 33%) and slope (10%).

The specific catchment-size ranges were modified slightly from BRT output to maintain sample size and the longitudinal effect of mean distribution, maintaining ecological conformance with RCC. Six final type/longitudinal groups were labeled by stream type and upper range of catchment area (km²) as: LS<1000, FS<40, FS<150, FS<550, FS<6000, FS>6000 hereafter referred to as drainage area groups (DAGs; Figure 7).

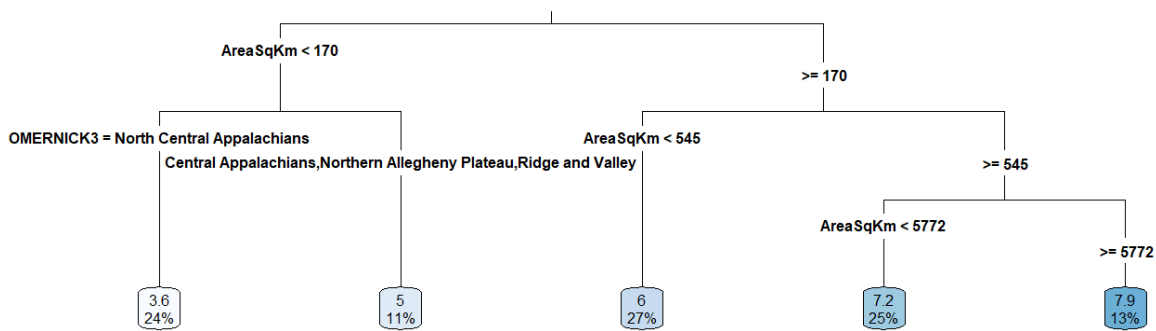


Figure 5. Boosted regression tree model of least disturbed sites showing important variables to classify freestone streams (FS) are generally related to catchment size and ecoregion. The bottom “leaflets” correspond to the mean thermal fish index (TFI) and the percentage of the dataset within each group.

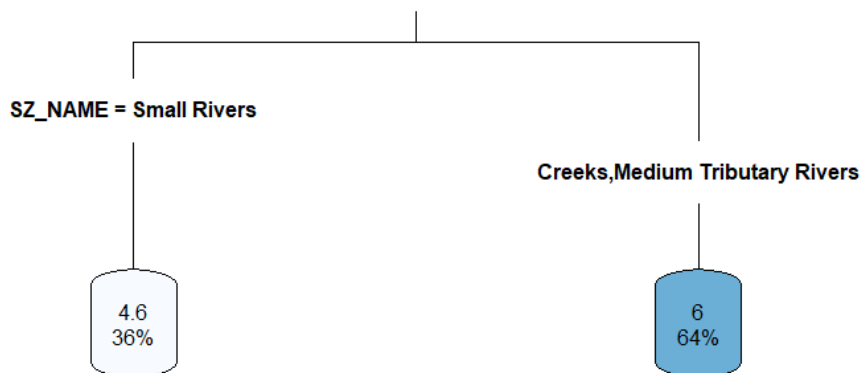


Figure 6. Boosted regression tree model showing important variables to classify limestone streams (LS), is generally related to stream size. All stress groups within the dataset were used as the sample size using only least disturbed sites precluded analysis.

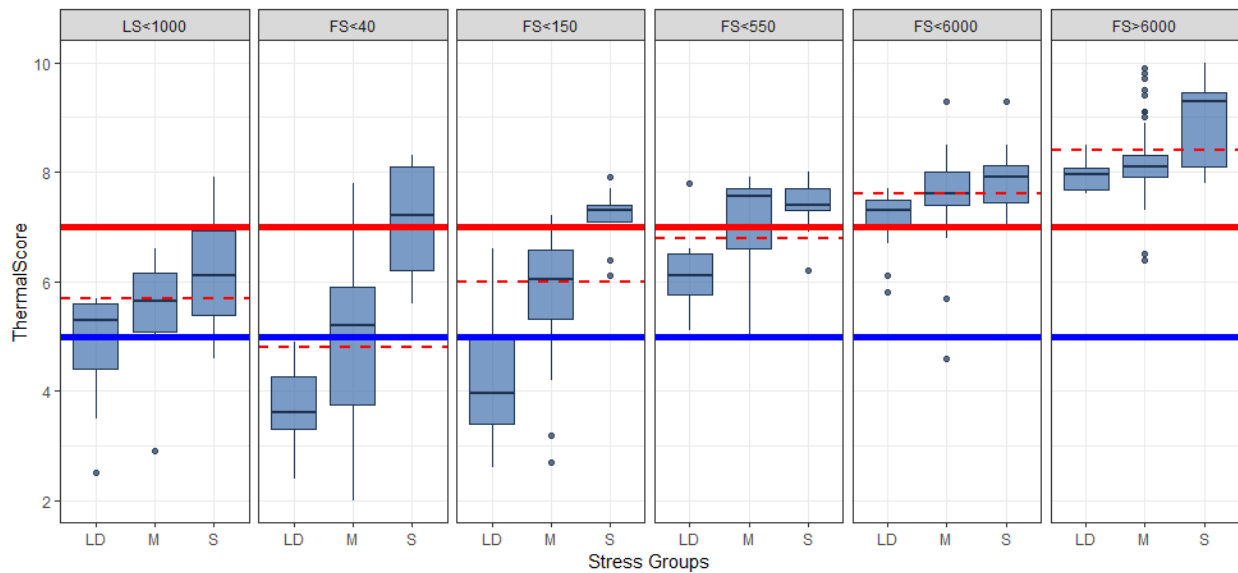


Figure 7. Boxplot of the final limestone (LS) and freestone (FS) drainage area groups (DAGs) (upper km² range). Stress groups are denoted as; Least Disturbed (LD), Moderate (M) and Stressed (S). Dotted red lines represents the 95th percentile of least disturbed sites signifying the impairment threshold. The solid blue line represents the upper limit for cold water assemblage and the solid red line represents the lower limit for warm water assemblage, transitional assemblage range is between.

Between-DAG comparisons of LD sites in FS streams using regression showed a significant increase in TFI score along a longitudinal gradient (adjusted $R^2 = 0.76$, $P = < 0.001$). All mean (and 95th percentile) TFI estimates were increasing as DAGs increased, maintaining ecological relevance. The 95th percentile for the LD sites within the LS DAG was 5.7. The 95th percentile for the LD sites within each longitudinally progressing FS DAG are as follows: 4.8, 6.0, 6.8, 7.6, 8.4. Discrimination efficiencies were all >80%, with the exception of the LS<1000 group that was 70%, averaging 88% across all DAGs (Table 6).

Table 6. Between-group and within-group results describing thermal scores across drainage area groups (DAG) and stress, respectively. Shared letters within the DAG column designate non-significant differences between width groups (Dunn’s test, $p < 0.05$). Kruskal-Wallis chi-squared test with bold italic represents significant results ($p < 0.01$). Sample sizes for each stress group and shared letters within the same cell designate non-significant differences in stress within width groups (Dunn’s test, $p < 0.05$).

DAG	n = LD, M, S	chi-squared	DE
LS<1000 ^{abc}	9 ^a , 6 ^{ab} , 10 ^b	4.92	70%
FS<40 ^{ab}	6 ^a , 31 ^a , 15	24.93	100%
FS<150 ^{ab}	16, 30, 9	27.65	100%
FS<550 ^{ac}	18, 30 ^a , 9 ^a	18.89	89%
FS<6000 ^{cd}	15, 30 ^a , 8 ^a	8.48	88%
FS>6000 ^d	6 ^a , 82 ^a , 11	8.57	82%
Average		15.57	88%

Precision estimates measured with CV and SD across all sites averaged 4.3% (TFI score ± 0.3) and 0.25 respectively. The highest CV and SD was noted in the FS<150 DAG averaging 8.8% (TFI score ± 0.7) and 0.4 respectively (Table 7). Classification efficiency, calculated to validate the calibration dataset and averaged across site width groups, was 95% for LD sites and 87% for stressed sites (

Table 8).

Table 7. Precision estimates using standard deviation (SD) and percent coefficient of variation (CV) for repeated sites within each drainage area group (DAG), regardless of stress level.

DAG	SD	CV %	N
LS<1000	0.2	4	16
FS<40	0.1	1.8	11
FS<150	0.4	8.8	59
FS<550	0.2	3.2	61
FS<6000	0.3	4.5	39
FS>6000	0.3	3.3	178

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Table 8. Validation classification efficiency, the percent stressed above impairment threshold and percent least disturbed under impairment threshold from validation dataset. Value in parenthesis denotes sample size.

	LS<1000	FS<40	FS<150	FS<550	FS<6000	FS>6000	Avg.
Least Disturbed	100% (2)	100% (2)	100% (4)	100% (4)	100% (6)	67% (3)	95%
Stressed	50% (4)	100% (1)	100% (3)	67% (3)	100% (1)	100% (3)	87%
Avg.	67%	100%	100%	86%	100%	83%	91%

Pairwise comparisons of the TFI to traditional metrics demonstrated numerous, significant correlations that were generally considered weak to moderate relationships (Figures 8-13). Strong and significant correlations were observed between the BCG5 and tolerant metrics throughout all DAGs. The omnivore metric tended to correlate with other metrics in larger streams but was considered highly variable. Relationships between the TFI and water quality were noted in all DAGs, except in the FS>6000 DAG, where the relationship was reduced. Across all DAGs the TFI consistently outperformed traditional metrics in response to water quality and habitat.

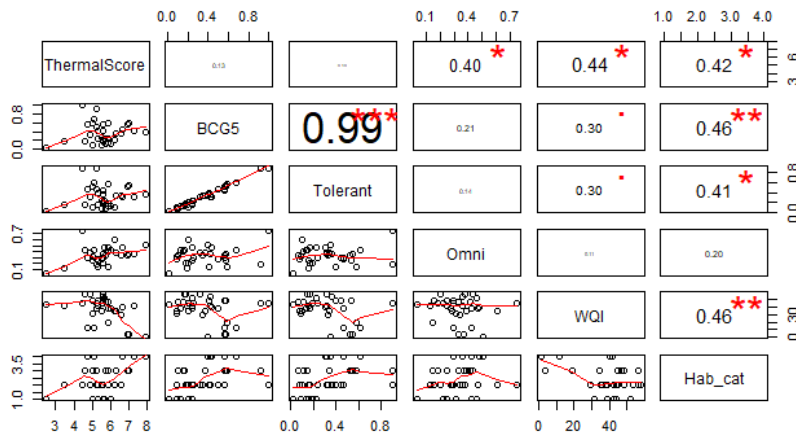


Figure 8. Pairwise comparison, using Spearman’s correlation coefficient, of the thermal fish index score (ThermalScore) to traditional metrics; Biological Condition Gradient category 5 (BCG5), percent tolerant, percent omnivorous (Omni), water quality (WQI) and habitat (Habcat) in LS<1000 streams. *** (P<0.001, ** (P<0.01), * (P<0.05)

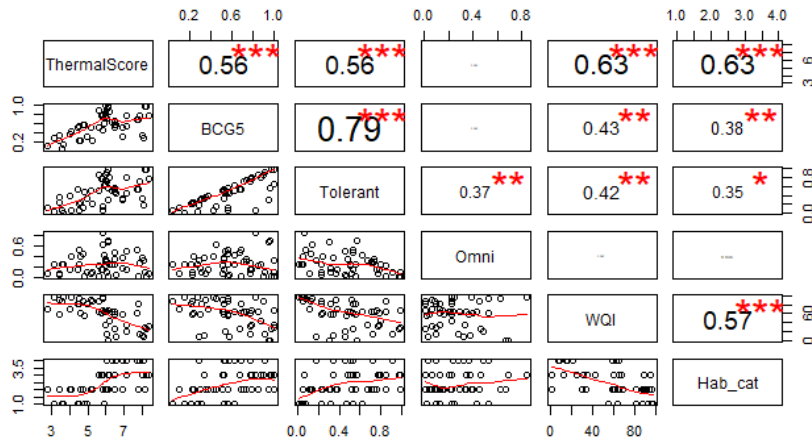


Figure 9. Pairwise comparison, using Spearman's correlation coefficient, of the thermal fish index score (ThermalScore) to traditional metrics; Biological Condition Gradient category 5 (BCG5), percent tolerant, percent omnivorous (Omni), water quality (WQI) and habitat (Habcat) in FS<40 streams. *** (P<0.001, ** (P<0.01), * (P<0.05)

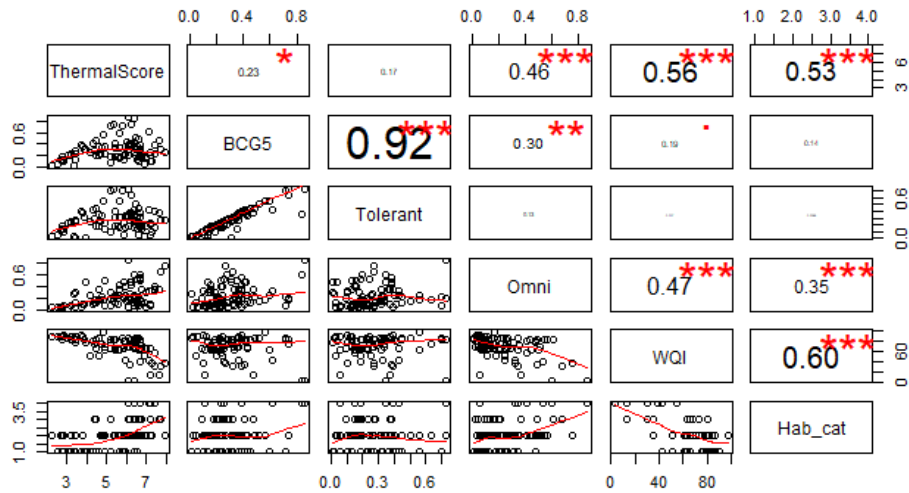


Figure 10. Pairwise comparison, using Spearman's correlation coefficient, of the thermal fish index score (ThermalScore) to traditional metrics; Biological Condition Gradient category 5 (BCG5), percent tolerant, percent omnivorous (Omni), water quality (WQI) and habitat (Habcat) in LS<1000 streams. *** (P<0.001, ** (P<0.01), * (P<0.05)

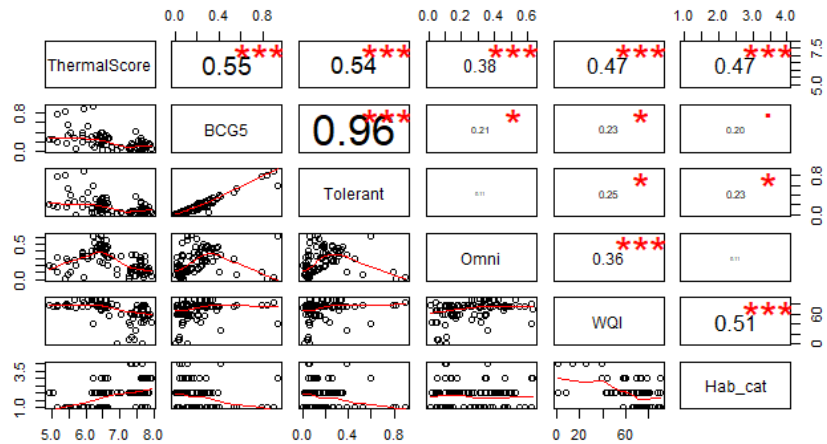


Figure 11. Pairwise comparison, using Spearman's correlation coefficient, of the thermal fish index score (ThermalScore) to traditional metrics; Biological Condition Gradient category 5 (BCG5), percent tolerant, percent omnivorous (Omni), water quality (WQI) and habitat (Habcat) in FS<550 streams. *** (P<0.001, ** (P<0.01), * (P<0.05)

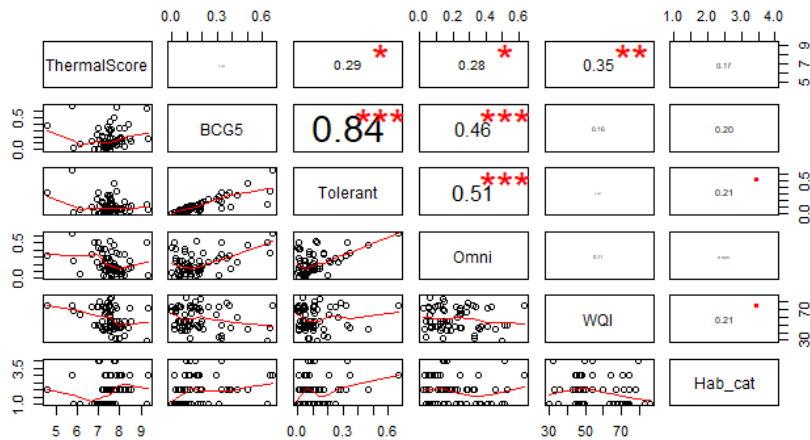


Figure 12. Pairwise comparison, using Spearman's correlation coefficient, of the thermal fish index score (ThermalScore) to traditional metrics; Biological Condition Gradient category 5 (BCG5), percent tolerant, percent omnivorous (Omni), water quality (WQI) and habitat (Habcat) in FS<6000 streams. *** (P<0.001, ** (P<0.01), * (P<0.05)

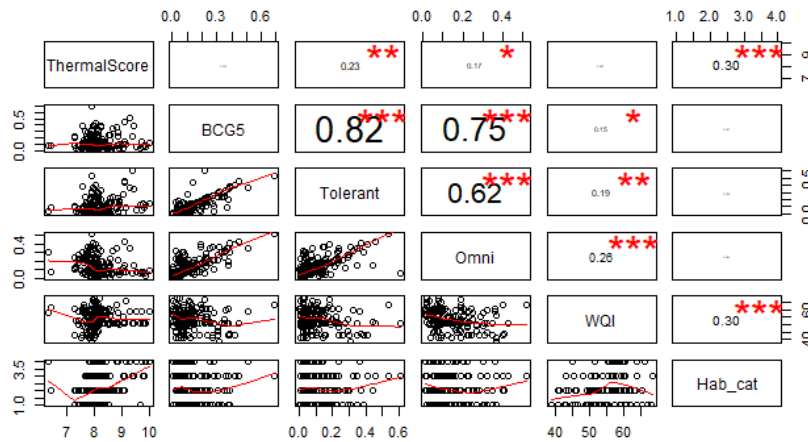


Figure 13. Pairwise comparison, using Spearman’s correlation coefficient, of the thermal fish index score (ThermalScore) to traditional metrics; Biological Condition Gradient category 5 (BCG5), percent tolerant, percent omnivorous (Omni), water quality (WQI) and habitat (Habcat) in FS>6000 streams. *** (P<0.001), ** (P<0.01), * (P<0.05)

DISCUSSION

Limestone streams were identified as a distinct group by their unique ability to support CWA in larger stream sizes than FS counterparts during preliminary classification investigations. Catchment area was determined to be the strongest predictor of the TFI in both FS and LS systems, with slope being an important (albeit secondary) predictor. This result is beneficial as it provides a template for the transition of fish assemblages along a longitudinal gradient for both stream types. The combination of these benefits allows for ecologically relevant, numerical estimates of expected thermal fish assemblages, based on environmental characteristics (e.g., DAGs). The longitudinal regression response of the TFI using the LD sites in this analysis was strong and significant (adjusted $R^2 = 0.76$, $P = < 0.001$) and meets expectations for ecological relevance, based on the RCC (Vannote et al. 1980).

On the surface, the LS<1000 DAG appears to be unique when compared to FS DAGs. This group had the lowest DE (70%) and lowest CE (in the S group; 50%) recorded. However, the reason for this apparent discrepancy is attributed to one major factor, trout-stocking. Herein, all (100%) LS streams that were considered S yet had a TFI score below the impairment threshold are streams regularly stocked with trout, thereby lowering the TFI score. This concept identifies a small degree of complexity that may be present in all fish-based bioassessments, where intentional (or unintentional) stocking co-occurs. Herein, a tradeoff exists between enhancing valuable recreational opportunities through stocking and measuring the response of fish assemblages that may not be solely driven by waterbody conditions. Subsequently, the effect of stocking

should be realized and treated as inherent, yet subtle “noise” that will likely be present in many fish-based bioassessments.

The lowest DE in the FS DAGs was noted in FS>6000, with a DE of 82%. This is attributed to; 1) a stress effect-size change and 2) using a four-tiered habitat category (Habcat) as a measure of stress. Generally, as stream size increases the range of water quality decreases; where large rivers tend to occupy a more-narrow and centralized distribution, as an effect of dilution (see generally, Nilsson 2008) (Figure 14). Alternatively, small streams are more susceptible to the extreme ends of the water quality range, being “very good” in heavily forested headwaters to “very poor” in effluent dominated headwaters. This important concept suggests as streams increase in size the effect of water quality stress may be mitigated, to some degree, and the effect of habitat quality may become more important. Using Habcat as a measure of habitat successfully standardized habitat stress across all methodologies employed, yet not without consequence. The Habcat is generally, but not always, comparable across all sites within the same stress level. For example, a river characterized as a 4 for sediment deposition may not be the same stress as a river that is impounded for miles, also characterized as a 4. This concept was the major driver of the reduced DE in FS>6000, where naturally occurring increases in sedimentation caused a site to fall in the S group. This reaffirms aforementioned confidence in correctly identified LD sites and reduced confidence in correctly identifying S sites.

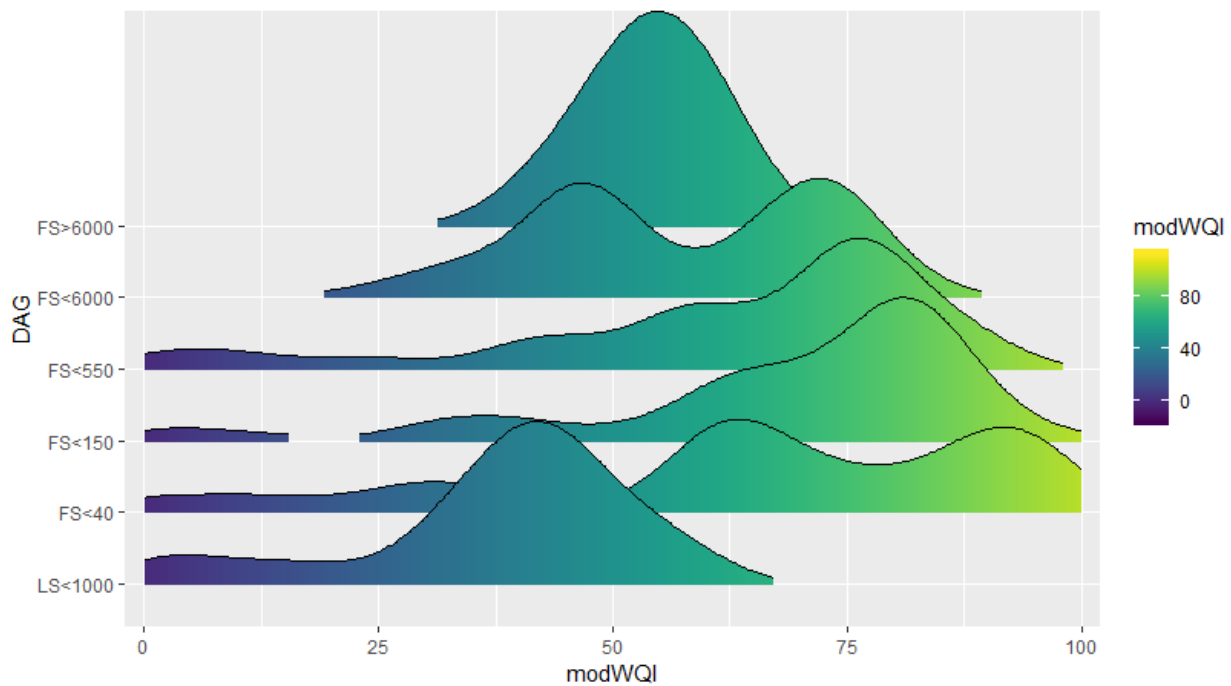


Figure 14. The shift in water quality range distribution across drainage area groups (DAG), as measured from the modified water quality index (modWQI).

The TFI was responsive to changes along a longitudinal gradient, temperature and stress (both habitat and water quality). The effect of water quality stress on the TFI was reduced longitudinally, as larger DAGs tended to occupy a narrow and more-central range of the modWQI (Figure 14). The effect of habitat on the TFI was important across both FS and LS groups (and DAGs) and tended to increase dramatically with increased sedimentation and impounding (Figure 15). These observations are important as the effects of multiple stressors are synergistic, antagonistic, or additive to the TFI scores. For example, as water quality is reduced by agricultural activities and loss of riparian areas, changes to instream habitat and temperature will likely parallel, having a dramatic effect on the TFI. Alternatively, a stream with mining influences may have reduced water quality, without drastic changes in habitat and temperatures, which may have a smaller effect on the TFI. In other words, as the number of stressors and/or intensity of stressors increases, increases in the TFI are expected. This is a desired outcome from a management perspective, as measured improvements in individual stressors may result in measurable recovery. For example, best management practices applied to small reaches of a larger watershed may have localized, measurable biological effects

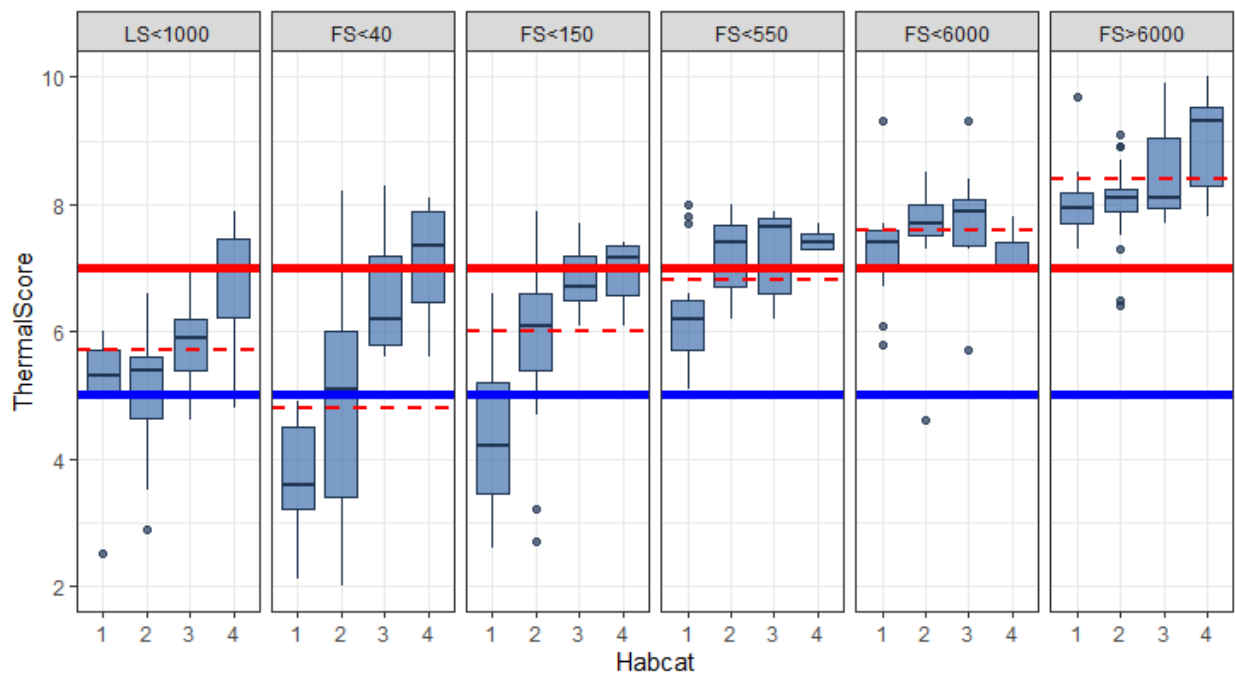


Figure 15. Boxplot of the final limestone (LS) and freestone (FS) drainage area groups (DAGs) (upper km² range). Habcat groups 1-4 are on a gradient of good to poor respectively. Dotted red lines represents the 95th percentile of least disturbed sites signifying the impairment threshold. The solid blue line represents the upper limit for cold water assemblage and the solid red line represents the lower limit for warm water assemblage, transitional assemblage range is between.

From a comparative perspective, the TFI may appear to be quite simple in design. In reality, the TFI should be viewed as a comprehensive metric, in that: 1) all species and individuals within the assemblage are provided equal consideration based on relative abundance, 2) can be applied uniformly across the State, basins, or ecoregions, 3) has an ecologically meaningful output of assemblage thermal class (cold vs. warm; as opposed to a purely statistically-derived construct), and 4) thermal preferences exhibit collinearity with other tolerances (water quality and habitat as evidenced herein) and traits. The TFI performed as well as or better than the three traditional metrics, in response to water quality and habitat conditions (Figures 8-13). The longitudinal stress effect-size change was again noted; where water quality and habitat elicited TFI responses in smaller DAGs and habitat became more important in the largest DAG (FS>6000), likely resulting from a more-central distribution along the water quality range (Figures 8-15).

Numerous unique samples were noted within the dataset that warrant further discussion. Unique samples within each DAG were apparent in both directions. The assemblages with lower TFI scores than the rest of the distribution were generally caused by: 1) hydrologic alterations in the form of augmented bottom-releases from upstream impoundments, 2) unique natural features such as increased groundwater volume or canopy cover, and 3) unrepresentative sample locations that are influenced strongly by proximal tributaries. Individual streams, or stream segments that have a natural ability to maintain colder fish assemblages can be viewed as unique and important from an ecological and/or recreational perspective. For example, the Delaware River near Balls Eddy, PA is in the FS<6000 DAG and has achieved TFI scores as low as 4.6. This exceptionally low score for such a large DAG is the result of flow management and cold water releases from upstream reservoirs. This portion of the Delaware river remains an important recreational destination for trout fishing. Conversely, while flow management and cold water augmentation scenarios may initially be portrayed as an improvement, it is not without consequences. An example of these consequences is apparent in Clarks Creek near Harrisburg, PA, a small tributary to the Susquehanna River. This stream is impounded by a drinking water reservoir; two fish survey sites were conducted, bracketing the reservoir. The site downstream of the reservoir had a catchment area of 62 km² (FS<150) with a TFI score of 5.4. This site is augmented by both cold water releases from the reservoir and trout stocking. The TFI score of 5.4 is within expected range based on its DAG; water quality is supportive of trout stocking and the assemblage is characterized as a TSA. The site upstream of the reservoir had a much smaller catchment area of 34km² (FS<40) with a TFI score of 7.8. The TFI score for this DAG is well above the 95th percentile of reference for the FS<40, set at 4.8. The upper site had excellent water quality and habitat but was located only 500 meters upstream of the impounded portion of the reservoir. Herein, the upstream site was influenced from fishes migrating upstream of the reservoir and was dominated

by the family Centrarchidae. In other words, fishes indigenous to a cold water habitat were being replaced by fishes indigenous to a warm water habitat; conceptually, a “thermally invasive species”. This effect is therefore considered a “biological pollution” as a result of significant habitat alterations within proximity (Pringle 1997, Elliott 2003). Consequently, beneficial changes to CWAs, downstream of cold water releases from impoundments was observed; but a reduction of CWAs within and upstream of impounded areas was also observed. Additionally, this tradeoff is likely to be in both directions of top release “spillover” impoundments (warmer assemblages upstream and downstream).

Overall, the TFI responded significantly to changes in longitude and stress in both freestone and limestone waterbodies across Pennsylvania. The discrimination and classification efficiencies were within acceptable range, averaging 88% and 91% across all groups respectively. Precision estimates measured from coefficient of variation were within (below) recommended threshold ranges of 10-15% (Stribling et al. 2008) and averaged 4.3%, with maxima still within acceptable limits at 8.8%. The TFI correlated with, and often outperformed traditional metrics in comparative analysis. These factors combined with added benefits of a large spatial application and ecological relevance solidify the TFI as a tool for assessing and evaluating fish assemblages across Pennsylvania’s lotic waterbodies.

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APPENDIX A – THERMAL PREFERENCE BY SPECIES

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Family	Common Name	Scientific Name	Numeric Value	Preference
Petromyzontidae	Ohio Lamprey	<i>Ichthyomyzon bdellium</i>	3	Cool
	Northern Brook			
Petromyzontidae	Lamprey	<i>Ichthyomyzon fossor</i>	3	Cool
Petromyzontidae	Silver Lamprey	<i>Ichthyomyzon unicuspis</i>	3	Cool
	American Brook			
Petromyzontidae	Lamprey	<i>Lampetra appendix</i>	3	Cool
Petromyzontidae	Sea Lamprey	<i>Petromyzon marinus</i>	3	Cool
Polydontidae	Paddlefish	<i>Polyodon spathula</i>	5	Warm
Lepisosteidae	Spotted Gar	<i>Lepisosteus oculatus</i>	5	Warm
Lepisosteidae	Longnose Gar	<i>Lepisosteus osseus</i>	5	Warm
		<i>Lepisosteus</i>		
Lepisosteidae	Shortnose Gar	<i>platostomus</i>	5	Warm
Amiidae	Bowfin	<i>Amia calva</i>	5	Warm
Hiodontidae	Goldeye	<i>Hiodon alosoides</i>	5	Warm
Hiodontidae	Mooneye	<i>Hiodon tergisus</i>	5	Warm
Anguillidae	American Eel	<i>Anguilla rostrata</i>	3	Cool
Clupeidae	Blueback Herring	<i>Alosa aestivalis</i>	5	Warm
Clupeidae	Skipjack Herring	<i>Alosa chrysochloris</i>	5	Warm
Clupeidae	Hickory Shad	<i>Alosa mediocris</i>	5	Warm
Clupeidae	Alewife	<i>Alosa pseudoharengus</i>	4	Cool-Warm
Clupeidae	American Shad	<i>Alosa sapidissima</i>	5	Warm
Clupeidae	Gizzard Shad	<i>Dorosoma cepedianum</i>	5	Warm
		<i>Campostoma</i>		
Cyprinidae	Central Stoneroller	<i>anomalum</i>	4	Cool-Warm
Cyprinidae	Goldfish	<i>Carassius auratus</i>	5	Warm
	Northern Redbelly			
Cyprinidae	Dace	<i>Chrosomus eos</i>	4	Cool-Warm
Cyprinidae	Finescale Dace	<i>Chrosomus neogaeus</i>	3	Cool
Cyprinidae	Redside Dace	<i>Clinostomus elongatus</i>	3	Cool
		<i>Clinostomus</i>		
Cyprinidae	Rosyside Dace	<i>funduloides</i>	2	Cold-Cool
		<i>Ctenopharyngodon</i>		
Cyprinidae	Grass Carp	<i>idella</i>	5	Warm
Cyprinidae	Satinfin Shiner	<i>Cyprinella analostana</i>	5	Warm
Cyprinidae	Spotfin Shiner	<i>Cyprinella spiloptera</i>	5	Warm
Cyprinidae	Common Carp	<i>Cyprinus carpio</i>	5	Warm
Cyprinidae	Streamline Chub	<i>Erimystax dissimilis</i>	4	Cool-Warm
Cyprinidae	Gravel Chub	<i>Erimystax x-punctatus</i>	3	Cool
Cyprinidae	Tonguetied Minnow	<i>Exoglossum laurae</i>	3	Cool
		<i>Exoglossum</i>		
Cyprinidae	Cutlip Minnow	<i>maxillingua</i>	3	Cool

Cyprinidae	Eastern Silvery Minnow	<i>Hybognathus regius</i>	5	Warm
Cyprinidae	Striped Shiner	<i>Luxilus chrysocephalus</i>	4	Cool-Warm
Cyprinidae	Common Shiner	<i>Luxilus cornutus</i>	4	Cool-Warm
Cyprinidae	Redfin Shiner	<i>Lythrurus umbratilis</i>	4	Cool-Warm
		<i>Macrhybopsis</i>		
Cyprinidae	Silver Chub	<i>storeriana</i>	4	Cool-Warm
Cyprinidae	Pearl Dace	<i>Margariscus margarita</i>	2	Cold-Cool
Cyprinidae	Hornyhead Chub	<i>Nocomis biguttatus</i>	3	Cool
Cyprinidae	River Chub	<i>Nocomis micropogon</i>	3	Cool
		<i>Notemigonus</i>		
Cyprinidae	Golden Shiner	<i>crysoleucas</i>	5	Warm
Cyprinidae	Comely Shiner	<i>Notropis amoenus</i>	5	Warm
Cyprinidae	Emerald Shiner	<i>Notropis atherinoides</i>	5	Warm
Cyprinidae	Silverjaw Minnow	<i>Notropis buccatus</i>	3	Cool
Cyprinidae	Blackchin Shiner	<i>Notropis heterodon</i>	4	Cool-Warm
Cyprinidae	Blacknose Shiner	<i>Notropis heterolepis</i>	4	Cool-Warm
Cyprinidae	Spottail Shiner	<i>Notropis hudsonius</i>	4	Cool-Warm
Cyprinidae	Silver Shiner	<i>Notropis photogenis</i>	4	Cool-Warm
Cyprinidae	Swallowtail Shiner	<i>Notropis procne</i>	3	Cool
Cyprinidae	Rosyface Shiner	<i>Notropis rubellus</i>	4	Cool-Warm
Cyprinidae	Sand Shiner	<i>Notropis stramineus</i>	4	Cool-Warm
Cyprinidae	Mimic Shiner	<i>Notropis volucellus</i>	4	Cool-Warm
Cyprinidae	Bluntnose Minnow	<i>Pimephales notatus</i>	4	Cool-Warm
Cyprinidae	Fathead Minnow	<i>Pimephales promelas</i>	4	Cool-Warm
	Eastern Blacknose			
Cyprinidae	Dace	<i>Rhinichthys atratulus</i>	3	Cool
Cyprinidae	Longnose Dace	<i>Rhinichthys cataractae</i>	3	Cool
	Western Blacknose			
Cyprinidae	Dace	<i>Rhinichthys obtusus</i>	3	Cool
		<i>Semotilus</i>		
Cyprinidae	Creek Chub	<i>atromaculatus</i>	3	Cool
Cyprinidae	Fallfish	<i>Semotilus corporalis</i>	4	Cool-Warm
Catostomidae	River Carpsucker	<i>Carpionodes carpio</i>	5	Warm
Catostomidae	Quillback	<i>Carpionodes cyprinus</i>	5	Warm
Catostomidae	Highfin Carpsucker	<i>Carpionodes velifer</i>	5	Warm
		<i>Catostomus</i>		
Catostomidae	Longnose Sucker	<i>catostomus</i>	2	Cold-Cool
		<i>Catostomus</i>		
Catostomidae	White Sucker	<i>commersonii</i>	3	Cool
Catostomidae	Creek Chubsucker	<i>Erimyzon oblongus</i>	4	Cool-Warm
Catostomidae	Northern Hog Sucker	<i>Hypentelium nigricans</i>	3	Cool
Catostomidae	Smallmouth Buffalo	<i>Ictiobus bubalus</i>	5	Warm
Catostomidae	Bigmouth Buffalo	<i>Ictiobus cyprinellus</i>	5	Warm

Catostomidae	Silver Redhorse	<i>Moxostoma anisurum</i>	4	Cool-Warm
Catostomidae	Smallmouth Redhorse	<i>Moxostoma breviceps</i>	4	Cool-Warm
Catostomidae	River Redhorse	<i>Moxostoma carinatum</i>	4	Cool-Warm
Catostomidae	Black Redhorse	<i>Moxostoma duquesnei</i>	4	Cool-Warm
Catostomidae	Golden Redhorse	<i>Moxostoma erythrurum</i> <i>Moxostoma</i>	4	Cool-Warm
Catostomidae	Shorthead Redhorse	<i>macrolepidotum</i>	4	Cool-Warm
Ictaluridae	Black Bullhead	<i>Ameiurus melas</i>	5	Warm
Ictaluridae	Yellow Bullhead	<i>Ameiurus natalis</i>	4	Cool-Warm
Ictaluridae	Brown Bullhead	<i>Ameiurus nebulosus</i>	4	Cool-Warm
Ictaluridae	Channel Catfish	<i>Ictalurus punctatus</i>	5	Warm
Ictaluridae	Stonecat	<i>Noturus flavus</i>	4	Cool-Warm
Ictaluridae	Tadpole Madtom	<i>Noturus gyrinus</i>	5	Warm
Ictaluridae	Margined Madtom	<i>Noturus insignis</i>	4	Cool-Warm
Ictaluridae	Brindled Madtom	<i>Noturus miurus</i>	4	Cool-Warm
Ictaluridae	Flathead Catfish	<i>Pylodictis olivaris</i>	5	Warm
Osmeridae	Rainbow Smelt	<i>Osmerus mordax</i>	1	Cold
Salmonidae	Cisco	<i>Coregonus artedi</i> <i>Coregonus</i>	1	Cold
Salmonidae	Lake Whitefish	<i>clupeaformis</i> <i>Oncorhynchus</i>	1	Cold
Salmonidae	Pink Salmon	<i>gorbuscha</i>	1	Cold
Salmonidae	Coho Salmon	<i>Oncorhynchus kisutch</i> <i>Oncorhynchus mykiss</i>	1	Cold
Salmonidae	Hybrid Golden Trout	(<i>hybrid</i>)	1	Cold
Salmonidae	Rainbow Trout	<i>Oncorhynchus mykiss</i> <i>Oncorhynchus</i>	1	Cold
Salmonidae	Steelhead	<i>mykiss(steelhead)</i> <i>Oncorhynchus</i>	1	Cold
Salmonidae	Chinook Salmon	<i>tshawytscha</i>	1	Cold
Salmonidae	Brown Trout	<i>Salmo trutta</i> <i>Salvelinus fontinalis x</i>	2	Cold-Cool
Salmonidae	Hybrid Tiger Trout	<i>Salmo trutta</i>	1	Cold
Salmonidae	Brook Trout	<i>Salvelinus fontinalis</i>	1	Cold
Salmonidae	Lake Trout	<i>Salvelinus namaycush</i> <i>Esox americanus</i>	1	Cold
Esocidae	Redfin Pickerel	<i>americanus</i> <i>Esox americanus</i>	4	Cool-Warm
Esocidae	Grass Pickerel	<i>vermiculatus</i>	4	Cool-Warm
Esocidae	Northern Pike	<i>Esox lucius</i>	4	Cool-Warm
Esocidae	Muskellunge	<i>Esox masquinongy</i>	4	Cool-Warm
Esocidae	Chain Pickerel	<i>Esox niger</i>	4	Cool-Warm
Umbridae	Central Mudminnow	<i>Umbra limi</i>	4	Cool-Warm

Percopsidae	Trout Perch	<i>Percopsis omiscomaycus</i>	1	Cold
Gadidae	Burbot	<i>Lota lota</i>	2	Cold-Cool
Atherinidae	Brook Silverside	<i>Labidesthes sicculus</i>	5	Warm
Fundulidae	Eastern Banded Killifish	<i>Fundulus diaphanus diaphanus</i>	5	Warm
	Western Banded Killifish	<i>Fundulus diaphanus menoma</i>	5	Warm
Fundulidae	Mummichog	<i>Fundulus heteroclitus</i>	5	Warm
Poeciliidae	Eastern Mosquitofish	<i>Gambusia holbrooki</i>	5	Warm
Belonidae	Atlantic Needlefish	<i>Strongylura marina</i>	5	Warm
Gasterosteidae	Fourspine Stickleback	<i>Apeltes quadracus</i>	1	Cold
Gasterosteidae	Brook Stickleback	<i>Culaea inconstans</i>	3	Cool
Gasterosteidae	Threespine Stickleback	<i>Gasterosteus aculeatus</i>	1	Cold
	Blackspotted Stickleback	<i>Gasterosteus wheatlandi</i>	1	Cold
Gasterosteidae	Ninespine Stickleback	<i>Pungitius pungitius</i>	1	Cold
Cottidae	Mottled Sculpin	<i>Cottus bairdii</i>	1	Cold
Cottidae	Blue Ridge Sculpin	<i>Cottus caeruleomentum</i>	1	Cold
Cottidae	Slimy Sculpin	<i>Cottus cognatus</i>	1	Cold
Cottidae	Potomac Sculpin	<i>Cottus girardi</i>	2	Cold-Cool
Cottidae	Spoonhead Sculpin	<i>Cottus ricei</i>	1	Cold
		<i>Myoxocephalus thompsoni</i>	1	Cold
Cottidae	Deepwater Sculpin	<i>Unidentified Cottus</i>	1	Cold
Moronidae	White Perch	<i>Morone americana</i>	5	Warm
Moronidae	White Bass	<i>Morone chrysops</i>	5	Warm
		<i>Morone chrysops x saxatilis</i>	4	Cool-Warm
Moronidae	Striped Bass	<i>Morone saxatilis</i>	4	Cool-Warm
Centrarchidae	Rock Bass	<i>Ambloplites rupestris</i>	4	Cool-Warm
Centrarchidae	Redbreast Sunfish	<i>Lepomis auritus</i>	4	Cool-Warm
Centrarchidae	Green Sunfish	<i>Lepomis cyanellus</i>	5	Warm
Centrarchidae	Pumpkinseed	<i>Lepomis gibbosus</i>	4	Cool-Warm
Centrarchidae	Warmouth	<i>Lepomis gulosus</i>	5	Warm
Centrarchidae	Orangespotted Sunfish	<i>Lepomis humilis</i>	5	Warm
Centrarchidae	Bluegill	<i>Lepomis macrochirus</i>	5	Warm
Centrarchidae	Longear Sunfish	<i>Lepomis megalotis</i>	4	Cool-Warm
Centrarchidae	Redear Sunfish	<i>Lepomis microlophus</i>	5	Warm
Centrarchidae	Smallmouth Bass	<i>Micropterus dolomieu</i>	4	Cool-Warm
Centrarchidae	Spotted Bass	<i>Micropterus punctulatus</i>	4	Cool-Warm
Centrarchidae	Largemouth Bass	<i>Micropterus salmoides</i>	5	Warm
Centrarchidae	White Crappie	<i>Pomoxis annularis</i>	5	Warm

		<i>Pomoxis</i>		
Centrarchidae	Black Crappie	<i>nigromaculatus</i>	5	Warm
Percidae	Greenside Darter	<i>Etheostoma blennioides</i>	4	Cool-Warm
Percidae	Rainbow Darter	<i>Etheostoma caeruleum</i>	4	Cool-Warm
Percidae	Iowa Darter	<i>Etheostoma exile</i>	3	Cool
Percidae	Fantail Darter	<i>Etheostoma flabellare</i>	3	Cool
Percidae	Johnny Darter	<i>Etheostoma nigrum</i>	4	Cool-Warm
Percidae	Tessellated Darter	<i>Etheostoma olmstedi</i>	3	Cool
Percidae	Tippecanoe Darter	<i>Etheostoma tippecanoe</i>	3	Cool
Percidae	Variagate Darter	<i>Etheostoma variatum</i>	3	Cool
Percidae	Banded Darter	<i>Etheostoma zonale</i>	4	Cool-Warm
		<i>Gymnocephalus</i>		
Percidae	Ruffe	<i>cernuus</i>	3	Cool
Percidae	Yellow Perch	<i>Perca flavescens</i>	3	Cool
	Chesapeake			
Percidae	Logperch	<i>Percina bimaculata</i>	4	Cool-Warm
Percidae	Logperch	<i>Percina caprodes</i>	4	Cool-Warm
Percidae	Channel Darter	<i>Percina copelandi</i>	4	Cool-Warm
Percidae	Gilt Darter	<i>Percina evides</i>	3	Cool
Percidae	Longhead Darter	<i>Percina macrocephala</i>	4	Cool-Warm
Percidae	Blackside Darter	<i>Percina maculata</i>	3	Cool
Percidae	Shield Darter	<i>Percina peltata</i>	3	Cool
Percidae	River Darter	<i>Percina shumardi</i>	4	Cool-Warm
Percidae	Sauger	<i>Sander canadensis</i>	3	Cool
		<i>Sander canadensis x</i>		
Percidae	Saugeye	<i>vitreus</i>	3	Cool
Percidae	Walleye	<i>Sander vitreus</i>	3	Cool
Sciaenidae	Freshwater Drum	<i>Aplodinotus grunniens</i>	5	Warm
		<i>Neogobius</i>		
Gobiidae	Round Goby	<i>melanostomus</i>	3	Cool